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# MITOCHONDRIAL DNA DIVERSITY IN FOUR POPULATIONS OF INDONESIAN FRESHWATER GIANT PRAWN (*Macrobrachium rosenbergii*)

### Keragaman DNA Mitokondria pada Empat Populasi Udang Galah (*Macrobrachium rosenbergii*) asal Indonesia

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#### ABSTRACT

Mitochondrial DNA has been widely applied to analyze fish genetic diversity, especially the cytochrome oxidase subunit I (COI) gene, which can be used as a marker of typical variation patterns, both intraspecific and interspecific crossbreeding. Haplotype diversity was observed in four populations of giant freshwater prawns (Macrobrachium rosenbergii) originating from Peureulak River in Aceh, Tabuk River in South Kalimantan, Bengawan Solo River in East Java, and the Siratu strain. Haplotype diversity was observed in the mitochondrial DNA COI gene of 1516 bp and restricted using 5 enzymes, namely Avall, Mspl, Rsal, HaelII, and Taql. The diversity of mtDNA COI haplotypes in the population of Peureulak River, Tabuk River, Bengawan Solo River, and Siratu were 0.8183, 0.6222, 0.7461, and 0.8044, respectively. The dendrogram of genetic distance showed that the Bengawan Solo River populations and the Siratu strain were in the same cluster, while Peureulak River and Tabuk River populations were in another group. The four populations can be used as genetic sources in crossbreeding activities based on haplotype data for each population, haplotype diversity, and genetic distance.

Keywords: COI, genetic distance, haplotype diversity, Macrobrachium rosenbergii, mtDNA

#### ABSTRAK

DNA Mitokondria telah banyak diaplikasikan untuk analisa keragaman genetik ikan, terutama gen cytochrome oxidase subunit I (COI) yang dapat digunakan sebagai penanda pola variasi yang khas baik pada persilangan intraspesifik maupun interspesifik. Keragaman haplotipe diamati pada empat populasi udang galah (*Macrobrachium rosenbergii*) yang berasal dari Sungai Peureulak di Aceh, Sungai Tabuk di Kalimantan Selatan, Sungai Bengawan Solo di Jawa Timur dan strain Siratu. Keragaman haplotipe diamati pada gen COI DNA mitokondria yang berukuran 1516 bp dan direstriksi menggunakan 5 enzim, yaitu Avall, Mspl, Rsal, HaeIII, dan Taql. Keragaman haplotipe mtDNA COI pada populasi Sungai Peureulak, Sungai Tabuk, Sungai Bengawan Solo dan Siratu masing-masing adalah 0,8183; 0,6222; 0,7461 dan 0,8044. Dendrogram jarak genetik menunjukkan bahwa populasi Sungai Bengawan Solo dan galur Siratu berada dalam 1 kluster yang sama, sedangkan populasi Sungai Peureulak dan Sungai Tabuk berada pada kluster lainnya. Berdasarkan data haplotipe untuk setiap populasi, keragaman haplotipe dan jarak genetik keempat populasi tersebut dapat dijadikan sebagai sumber genetik dalam kegiatan persilangan.

Kata Kunci: COI, jarak genetik, keragaman haplotipe, Macrobrachium rosenbergii, mtDNA

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#### INTRODUCTION

Giant freshwater prawn (Macrobrachium rosenbergii) is a species of freshwater shrimp that has the potential to be developed to support animal protein-based of national food self-reliance and security programs. Giant freshwater prawns are Indonesian freshwater fishery commodities with high economic value for domestic consumption and export. The market demand that can be met is still limited to domestic consumption, especially seafood in restaurants, high-class restaurants, and highstar hotels. Meanwhile, export demand from Japan, the United States and European countries, and Singapore has not been fulfilled

The development of giant freshwater prawn cultivation in Indonesia is relatively lower compared to other countries such as China, Vietnam. India, Thailand, and Bangladesh. Until this time, China is the number one producer of giant freshwater prawns, with a total production of 132,678 tons in 2016, while Indonesia is in 4th position after Bangladesh and Thailand, with a total output of 11,708 tons (Farook et al. 2019). productivity of Indonesian The giant freshwater prawns is still low, from 2010 to 2016, only increasing by 2.86% per year. Currently, the cultivation of giant freshwater prawns has developed in other countries such as the United States of America and China (Zafar et al. 2015). FAO (2020) reports that the global production of giant freshwater prawns has not shown a significant increase. Its production in 2018 reached 234,400 tons, only a rise of 7.67% compared to production in 2010. Furthermore, the production of giant freshwater prawns was only 2.5% of the total crustacean production.

In Indonesia, giant freshwater prawn seeds are currently being produced in hatcheries using wild catches for cultivation purposes. Domestication efforts and genetic improvement of giant prawns have been carried out. Giant freshwater prawns GMacro II and Siratu are two giant freshwater prawn strains produced by genetic improvement efforts released by the government in 2014 (KKP 2014) and 2015 (KKP 2015). Due to the limited availability of genetically improved giant freshwater prawns broodstock, the breeders make efforts to bring in natural broodstock from various locations to produce seeds. Crossbreeding between quality populations that are carried out usually comes from populations from different Crossbreeding deographies. between populations/ strains is common in fish farming to avoid inbreeding. Crossbreeding between controlled populations/ strains is expected to produce offspring that have better growth characteristics, are more resistant to disease, and have low levels of abnormalities (Tave 1993).

The crossbreeding process will be more effective if the genetic quality can be identified first. Genetic quality can be analyzed through the level of gene diversity because high genetic diversity in a population means that there are many individuals with a variety of different traits. Genetic diversity is significant for a population to adapt to a dynamic environment (Tave 1993). Aliah (2014) has carried out an analysis of genetic diversity on several tilapia strains (Oreochromis niloticus), which will be used to develop hybrid Salina tilapia resistant to high salinity. Then the same was done by Mastrochirico-Filho et al. (Piaractus fish (2019) on pacu mesopotamicus) to obtain a parent base population used in breeding activities for this species. Several studies on crosses between populations or strains that produce offspring that have better growth and survival have been reported by Aliah (2014) on Salina tilapia (Oreochromis sp.), Sunarma et al. (2016) on African catfish (Clarias gariepinus), Zeng at al. (2017) on mandarin fish (Siniperca chuatsi) and several strains of tilapia. O. niloticus (Novelo et al. 2021).

Several methods that can be used to analyze genetic diversity in fish include DNA microsatellites (Cordova-Alarcon et al. 2019), PASA (PCR amplification of specific alleles), and SNP (single nucleotide polymorphism) (Dudu et al. 2015). Genetic diversity can be estimated molecularly. One of them is by using genetic markers of mitochondrial DNA. The mitochondrial DNA control region (mtDNA CR) is the most variable part of the mtDNA, composed of a conservative central area with relatively divergent left and proper domains. It involves 3-5 times more rapidly than the other segments of the mitochondrial genome (Cheng et al. 2012, Li et al. 2022). In addition, the type of mutation in mtDNA is simple, namely base substitution or base

length mutation, and occurs mainly in small non-coding regions; thus, the mtDNA polymorphism is a neutral genetic marker. The COI (Cytochrome Oxidase I) region of mtDNA is generally applied in the genetic structure of the fish population (Ceruso et al. 2020) and reliable DNA barcodes for the identification and discrimination of fish species (Gao et al. 2020).

Mitochondrial DNA sequences or sequenced *mitogenomes* of giant freshwater prawns have been studied, including giant freshwater prawns from China with the size of 15,767 bp (Li et al. 2021), giant freshwater prawns from Indonesia with the size of 15,772 bp with the GenBank accession number of AY659990 (Ma et al. 2011) and giant freshwater prawns from China 15,766 bp (Li et al. 2019). Mitochondrial DNA sequences from several Macrobrachium genera, including М. nipponense, have а mitochondrial genome sequence of 15,806 bp (GeneBank ID NC\_015073.1), and M. lanchesteri has a shorter mitochondrial sequence of 15,694 bp with GeneBank ID NC\_0122217.1 (Ma et al. 2011). The mitochondrial DNA genetic markers have been successfully used to analyze differences in genetic diversity between natural and aquaculture populations (Nguyen Thanh et al. 2015), identify various types of tilapia species (Wu and Yang 2012), and evaluate the restocking program in fish species (Li et al. 2016, Maidin et al. 2017).

This study was conducted to identify the genetic diversity of four populations of giant freshwater prawns (*M. rosenbergii*) originating from the Peureulak River (Aceh), Tabuk River (South Kalimantan), Bengawan Solo River (East Java), and the Siratu (genetically improved strain), which will be used as parent base in the process of



Figure 1. Giant freshwater prawn, *M. rosenbergii* 

crossbreeding. The population was selected based on differences in the area's geographic location to obtain high potential for diversity, namely natural giant freshwater prawns from Sumatera Island, Kalimantan Island, Java Island, and genetically improved giant freshwater prawns. Genetic diversity was carried out using mitochondrial DNA COI gene markers. Hopefully, the genetic diversity data obtained can be used as primary data to determine parent combinations in the crossbreeding process.

#### MATERIALS AND METHODS

#### Location and time

Research on mitochondrial DNA diversity in four populations of Indonesian giant freshwater prawns (*M. rosenbergii*) was conducted at the Bioaquaculture Laboratory, Center for Agricultural Production Technology located at LAPTIAB BRIN, PUSPIPTEK, South Tangerang, Banten. This research was conducted from 2017-to 2018.

#### Materials

The 194 samples of caudal fin pieces were collected from 4 genetic sources of giant freshwater prawns (Figure 1), namely, 62 samples from the Peureulak River in Aceh (4°55'58" N 97°55'45" E, alt. 93 mamsl), 44 samples from the Bengawan Solo River in East Java (7°20'28" S 111°41'23" E, alt. 295 mamsl), 18 samples from the Tabuk River in South Kalimantan (3°17'49" S 114°42'14" E, alt. 37 mamsl) and 42 samples of giant freshwater prawns of Siratu strain from the Pelabuhan Ratu Prawn Hatchery Center (6°57'33" S 106°28'14" E, alt. 1.055 m mamsl). Giant freshwater prawns of the Siratu strain are the result of genetic improvement carried out by BBPBAT-KKP Sukabumi and have been released by the Minister of Fisheries and Marine Affairs in the Decree of the Minister of Fisheries and Marine Affairs Number 25/KEPMEN-KP/2015 on April 4, 2015 (KKP 2015).

The DNA extraction was done using the modified standard SDSphenol/chloroform method (Green and Sambrook 2012). The DNA was extracted from 50 mg pleopod of giant freshwater prawn samples. The 700  $\mu$ L of lysis solution (10 mM Tris-HCl pH 7.5, 1.5 M NaCl, 10 nM EDTA pH 7.5, 0.5% SDS, and 4 M Urea) and 5  $\mu$ L

Proteinase K (1 mg  $\mu$ L<sup>-1</sup>) were mixed and vortexed, before the incubation for 24 hours at 37°C. After the incubation, 700 µL of phenol/chloroform/isoamyl alcohol (25:24:1) and chloroform/isoamyl alcohol (24:1) were added. The supernatant was taken and added to 10% 3M sodium acetate and 2 times the volume of absolute ethanol, followed by incubation on ice for 30 minutes and centrifugation at 5000 rpm for 10 minutes. The pellet was added with 1 mL of 70% ethanol and centrifuged again at the same speed. The supernatant was discarded, and the pellet was dried. The pellet was dissolved in TE buffer (10 mM Tris-HCl pH 7.2; 1 mM EDTA pH 8) with a volume of 20-100 µL and stored at 4°C.

Cytochrome Oxidase I (COI) regions in mtDNA were selected to detect haplotype diversity. The D-loop regions were amplified using COI UD forward 5'-ACG CAA CGG TGG CTT TTC-3' primer (GenBank No. Acc. AY659990.1) and reverse 5'-TAG TTA GCT GTT AGG GGG AT-3' (Liu et al. 2007) with a target of 1516 bp. The PCR reaction proceeded as follows: pre-denaturation at 95°C for 5 minutes followed by 35 cycles (95°C for 1 minute, 56°C for 30 seconds, and 72°C for 1 minute). Furthermore, the diversity of COI mtDNA genes was analyzed using 5 restriction enzymes, namely Avall (G/GCC), Mspl (CC/GG), Rsal (GT/AC), Haelll



Figure 2. COI gene of mtDNA region in four female giant freshwater prawns from Peureulak River, Aceh (AB9-AB12) (1516 bp)

(GG/CC), and *Taq*I (T/CGA). The enzyme restriction reaction used a total volume of 10  $\mu$ L with the following composition: 1  $\mu$ L of sample DNA, 1 U of *Ava*II enzyme, 2  $\mu$ L of buffer O, and 6  $\mu$ L of nuclease-free water.

Analysis of mtDNA variation was carried out to detect genetic variation of giant freshwater prawns based on the presence or absence of restriction sites on mtDNA COI. Each restriction enzyme has a different restriction pattern, and each individual has a restriction pattern that may be different for each enzyme used. Based on the restriction pattern, a haplotype was made. Haplotype diversity (h) in a population is calculated according to Nei's (1978) equation:

$$h = 2n \left( \sum_{i=1} xi^2 \right) / (2n - 1)$$

where,

h: haplotype diversity

n: sample size

xi: haplotype frequency of sample-i

The genetic distance between populations is presented in a UPGMA matrix and dendrogram, which was analyzed from haplotype data using the TFPGA (tools for population genetic analyses) program ver 1.3.

### **RESULTS AND DISCUSSION**

The COI mtDNA gene from 166 giant freshwater prawn samples was amplified using primers COI UD forward (5'-ACG CAA CGG TGG CTT TTC-3') and COI UD reverse (5'-TAG TTA GCT GTT AGG GGG AT-3') with the size of 1,516 bp (Figure 2). Cytochrome Oxidase I (COI) region is one of the proteincoding for mtDNA. The COI gene is commonly used as a genetic marker in population studies. Species with very closely categorically genetic related. can be distinguished using mtDNA COI sequence differences at particular nucleotide positions (In et al. 2017).

The restriction type of Avall and Taql enzymes produced two patterns (Figure 3a), Haelll enzymes had three patterns (Figure 3b), Mspl enzymes had four patterns, and Rsal enzymes produced three patterns (Figure 3c). The results of the restriction of the COI mtDNA area using five enzymes Avall, Mspl, Rsal, Haelll, and Taql in four populations of giant freshwater prawns

produced 30 haplotypes. The giant freshwater prawn population from Peureulak River, Aceh has 15 haplotypes, the population from Tabuk River, South haplotypes, Kalimantan. has five the population from Bengawan Solo River has eight haplotypes, and the population strain Siratu has 10 haplotypes (Table 1). The AABAA haplotype was found in three observed populations, namely the population of Tabuk River, Peureulak River, and Bengawan Solo River, with the frequencies of 0.5556, 0.2742, and 0.0227, respectively. Two haplotypes, namely ABCBA and ABCCA, were found in Bengawan Solo River and Siratu populations. In comparison, two other haplotypes, AABBA and AABCA, were found in the populations of Peureulak River and Bengawan Solo River. Several haplotypes were found dominant in each population, namely AABAA in the Tabuk River population with the frequency of 0.5556, ABCAA in the Siratu population with the frequency of 0.3571, and ABCBA in the Bengawan Solo River population with the frequency of 0.4545. Furthermore, two haplotypes were found in the Peureulak River population, namely BABAB and AABAA, with the frequencies of 0.3065 and 0.2742, which were relatively high compared to other haplotypes in the population.

The diversity of haplotypes in a population is highly dependent on the number of detected haplotypes and the frequency of each haplotype. The haplotype diversity observed in the four giant freshwater prawn populations was 0.8183 in the Peureulak River population, 0.8044 in the Siratu

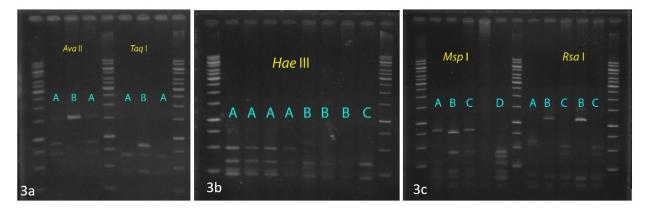


Figure 3. Restriction patterns of COI gene of mtDNA in 4 populations of giant freshwater prawn (*M. rosenbergii*) by using Avall and Taql (a), *Hae*III (b), *Msp*I and *Rsa*I (c) enzymes

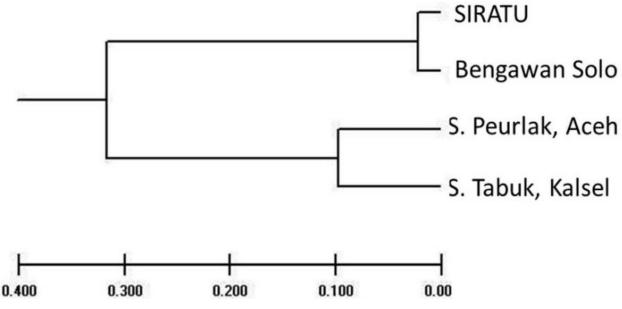


Figure 4. UPGMA dendrogram based on Nei's genetic distance among 4 populations giant freshwater prawn (*M. rosenbergii*) on COI mtDNA sequences

No.	Haplotype	S. Peurlak		B. Solo		S. Tabuk		SIRATU	
		n	frequency	n	frequency	n	frequency	n	frequency
1	AAAA	2	0.0323			1	0.0556		
2	AAABA							1	0.0238
3	AABAA	17	0.2742	1	0.0227	10	0.5556		
4	AABAB	5	0.0806						
5	AABAC					1	0.0556		
6	AABBA	3	0.0484	1	0.0227				
7	AABCA	1	0.0161	5	0.1136				
8	AABCB	2	0.0323						
9	AACAA	1	0.0161						
10	AACAB	1	0.0161						
11	AACBA			6	0.1364				
12	AACCA			5	0.1136				
13	AADCB	1	0.0161						
14	ABBAA					5	0.2778		
15	ABBBA			2	0.0455				
16	ABCAA							15	0.3571
17	ABCBA			20	0.4545			6	0.1429
18	ABCCA			4	0.0909			8	0.1905
19	ACCAA							1	0.0238
20	ACCBA							4	0.0952
21	ACCCA							1	0.0238
22	ADBAA	1	0.0161						
23	ADCBA							1	0.0238
24	BAAAA	1	0.0161						
25	BABAA	4	0.0645			1	0.0556		
26	BABAB	19	0.3065						
27	BABBA							4	0.0952
28	BABBB	3	0.0484						
29	BABCB	1	0.0161						
30	BBBBA							1	0.0238
No. of Sample		62		44		18		42	
	26BABAB27BABBA28BABBB29BABCB30BBBBANo. of SampleNo. of Haplotype		15		8		5		10
Haplot	type Diversity		0.8183		0.7461		0.6222		0.8044

 Table 1. Haplotype diversity in 4 populations of giant freshwater prawn (*M. rosenbergii*) based on haplotype frequency of mtDNA COI gen with 5 restriction enzymes (*Avall, Mspl, Rsal, Haelll, and Taql*)

population, 0.7461 in the Bengawan Solo River population, and 0.6222 in the Tabuk River giant freshwater prawn population (Table 2). Referring to the genotypic diversity index according to Nei (1978), namely 0.1-0.4 for low diversity, 0.5-0.7 for moderate diversity, and 0.8-1.0 for high diversity. The haplotype diversity of giant freshwater prawns from the Tabuk River was classified as moderate, while giant freshwater prawns from Peureulak River, Bengawan Solo River, and Siratu strains were classified as high. The lower haplotype diversity in the shrimp population from Tabuk River was probably caused by the smaller number of samples observed compared to the sample size of three other giant freshwater prawn populations.

The more similar haplotypes the giant freshwater prawn populations observed, the closer their kinship or the smaller genetic distance value. The genetic distance value in the form of a matrix among four giant freshwater prawn populations is presented in Table 2. The genetic distance between the giant freshwater prawn population of Bengawan Solo River and the Siratu strain is the closest at 0.0202, while the genetic 

 Table 2.
 Genetic distance between populations of giant freshwater prawns (M. rosenbergii) from Peureulak River (Aceh), Bengawan Solo River (East Java), Tabuk River (South Kalimantan) and Siratu

	Peureulak River	Bengawan Solo River	Tabuk River	Siratu
Peureulak River	*****			
Bengawan Solo River	0,3599	*****		
Tabuk River	0,0959	0,2766	*****	
Siratu	0,3603	0,0202	0,2641	*****

distance between the Peureulak River population and Siratu is the farthest with a value of 0.3603.

The genetic distance dendrogram presented in Figure 4 shows that the giant freshwater prawns from Bengawan Solo River and Siratu are closely related to form a cluster far apart from giant freshwater prawns derived from Peureulak River and Tabuk River. Furthermore, the genetic distance of the giant freshwater prawn Siratu – Bengawan Solo River was much smaller (0.0202) than the genetic distance between the giant freshwater prawn populations from Peureulak River and Tabuk River (0.0959).

In this study, mitochondrial DNA genetic markers have been carried out to identify and analyze the diversity of the giant freshwater prawn population, which will be used as the primary parent population in crossbreeding activities. Mitochondrial DNA genetic markers successfully examined the differences in genetic diversity of natural and aquaculture populations of giant freshwater prawns in China and Vietnam using partial sequences of the 16S rDNA gene and mitochondrial DNA COI gene (Nguyen Thanh et al. 2015). This method was also applied to identify various types of tilapia: the Oreochromis genus, Sarotherodon genus, Tilapia, and the hybrid O. massambicus and O. niloticus, which have been introduced to Hawaii (Wu and Yang 2012). Moreover, Zhang et al. (2013) found a decrease in the genetic diversity of sturgeon fish (Acipenser sp.) in nature due to the presence of hybrid sturgeon fish secondgeneration mass production in hatcheries released into the wild. Zhang et al. (2016) proved that mitochondrial DNA markers were inherited "maternally" in the hybridization species. process between 4 fish Megalobrama sp and Parabramis pekinensis first and second generations. Furthermore, mitochondrial DNA genetic markers were used to evaluate the restocking program on the endangered species *Percocypris pingi* in the Yalong River, China (Li et al. 2016). They assessed the success of restocking activities of giant freshwater prawns (*M. rosenbergii*) to the Petagas River in the Sabah region, Malaysia, between 2012 – 2015 (Maidin et al. 2017).

According to Nei (1978), the genetic prawn giant freshwater diversity of populations from the Peureulak River and Siratu strains is relatively high. This indicates that genetic improvement activities in giant freshwater prawns have been successful and can be used to produce good quality seeds of giant freshwater prawns as long as the parent management is carried out properly. Several comparisons of genetic diversity between natural and cultivated populations have been completed to anticipate the effect of inbreeding in cultured populations whose seeds are produced in hatcheries. Nguyen Thanh et al. (2015) analyzed the mtDNA diversity in cultured prawn populations from Zhejiang, Guangdong, and Guangxi provinces in China and natural two populations from the Mekong River and Dong Nai River in Vietnam. The results showed that wild populations had higher genetic diversity than cultured populations. Bala et al. (2017) also reported that the genetic diversity of the giant freshwater prawn population in Bangladesh of natural origin was higher than the genetic diversity of the giant freshwater prawn population in the hatcherv. Furthermore, Li et al. (2016) reported that the restocking activity of P. pingi fish species hatchery products in Moulo River, China, did not affect the level of genetic diversity of natural populations, which remained high and higher than the genetic diversity of hatchery populations, even though the number of fish species in the nature is much decreased due to anthropogenic factors.

On the other hand, the genetic diversity of giant freshwater prawn populations in four rivers in the Malaysia Peninsula has decreased its heterozygosity, and inbreeding occurs due to overfishing of giant freshwater prawns to produce seeds in hatcheries (Atin et al. 2017). Furthermore, Binur and Pancoro (2017) observed the diversity of four microsatellite DNA loci of Indonesia's giant freshwater prawn hatchery population. The results indicated inbreeding in the moderate category at the post-larval stage at 4 giant freshwater prawn hatchery locations on Java Island. Therefore, using post larvae from these four hatcheries is not recommended for potential broodstock. Different results were reported by Li et al. (2017); in spotted barbell fish (Hemibarbus maculates), cultured populations produced from controlled hatcheries have better genetic diversity than natural populations that are naturally selected due to the decrease in environmental quality.

The kinship pattern of a population occurs because of the spread and process of migration (gene flow), which moves genetic material from one population to another. If there are two populations with different genetic structures and then there is a transfer of genes between populations within a specific time, it will be genetically similar. The transfer of genes between populations can be caused by the movement of individuals or populations naturally due to the location of the habitats of these two populations being close together or due to human intervention.

In this study, the kinship pattern among four observed giant freshwater prawn populations could be analyzed from the resulting genetic distance. The Java Island cluster consisting of the Bengawan Solo River population and the Siratu strain had a lower genetic distance value than the outside of the Java Island. The Java Island cluster consists Peureulak River and Tabuk River of populations, with high genetic distance values between them, reflecting the absence of gene flow from these two natural populations. Furthermore, the results showed that not one haplotype detected even was simultaneously in the 4 populations.

This genetic diversity and distance observed in this study are valuable basic information for determining the combination of parental populations in cross-breeding to obtain offspring with the desired traits. The results of the analysis showed that the genetic diversity of the four giant freshwater prawn populations is relatively high (0.6222 – 0.8183) and has a relatively large genetic distance so that it can become the primary population for cross-breeding. The hybrid/crossbreed giant freshwater prawns are expected to be positive heterosis, which means they have better characteristics than their parents.

The National Research and Innovation Agency (known as BRIN) has initiated programs to improve the genetic quality of giant freshwater prawns, one of which is cross-breeding activities to obtain hybrids with superior characteristics. For this crossing activity, giant freshwater prawns have been obtained from three different geographic locations, plus the genetically improved giant freshwater prawn strains released by the government. Based on the results of the analysis of genetic diversity and genetic distance of the four observed populations of giant freshwater prawns, namely Peureulak River, Tabuk River, Bengawan Solo River, and Siratu, a preliminary trial of reciprocal crossbreeding between populations of (1) Peureulak River and Bengawan Solo River, (2) Bengawan Solo River and Siratu and (3) Peureulak River and Siratu. The results of rearing for 120 days showed that reciprocal crossbreeding of (1) Peureulak River and Bengawan Solo River at the juvenile stage had daily growth rates and better survival rates than reciprocal crossbreeding of (2) Bengawan Solo River and Siratu and (3) Peureulak River and Siratu.

### CONCLUSION

The diversity of mitochondrial DNA COI gene haplotypes observed in giant freshwater prawn populations from Peureulak River, Tabuk River, Bengawan Solo River, and Siratu strains showed relatively high diversity ranging from 0.6222 to 0.8183. The genetic distance between these four populations ranged from 0.0202 to 0.3603 and formed two clusters: the Bengawan Solo River population – the Siratu strain cluster, and the Peureulak River population – Tabuk River population cluster. Based on the haplotype diversity data and genetic distance, these four giant freshwater prawn populations can be used as genetic sources for crossbreeding activities.

## REFERENCES

- Aliah RS (2014) Development of SALINA -Saline Indonesian Tilapia. In Sudarvono A and Mufid A (ed), ICAI 2014 Proceedings. International Conference of Aquaculture Indonesia 2014. Toward a Better and Global Sustainable Industry. Bandung, Indonesia, June 20-21, 2014. Pp. 225 - 232. ISSN 2356 -0800
- Atin KH, Christianus A, Fatin N, Lutas AC, Shabanimofrad M, Subha B (2017) Genetic diversity analysis in Malaysian giant prawns using expressed sequence tag microsatellite markers for stock improvement program. Genet Mol Res 16: gmr16035685. doi: 10.4238/gmr16035685
- Bala B, Malik M, Saclain S, Islam MS (2017) Genetic variation in wild and hatchery populations of giant freshwater prawn (*Macrobrachium rosenbergii*) revealed by randomLy amplified polymorphic DNA markers. J Genet Eng Biotechnol 15:23-30. doi: 10.1016/j.jgeb. 2017.02.006
- Binur R, Pancoro A (2017) Inbreeding depression level of post-larvae freshwater prawn (*Macrobrachium rosenbergii*) from several hatcheries in Java, Indonesia. Biodiversitas 18:609-618. doi: 10.13057/biodiv/d180223
- Ceruso M, Mascolo C, De Luca P, Venuti I, Smaldone G, Biffali E, Anastasio A, Pepe T, Sardino P (2020) A rapid method for identification of fresh and processed *Pagellus erythrinus* species against frauds. Foods 9:1397. doi: 10.3390/foods9101397
- Cheng YZ, Xu TJ, Jin XX, Tang D, Wei T, Sun YY, Meng FQ, Shi G, Wang RX (2012) Universal primer for amplification of the complete mitochondrial control region in marine fish species. Mol Biol 46:810-813. doi: 10.1134/S0026893312040024
- Cordova-Alarcon VR, Araneda C, Jilberto F, Magnolfi P, Toledo MI, Lam N (2019) Genetic diversity and population structure of *Genypterus chilensis*, a commercial benthic marine species of

South Pacific. Front Mar Sci 6:748. doi: 10.3389/fmars.2019.00748

- Dudu A, Georgescu SE, Costache M (2015) Evaluation of genetic diversity in fish using molecular markers.Chapter 7. In: Caliskan M, Oz GC, Kavakli IH, Ozcan B (Eds). Molecular Approch to Genetic Diversity, Intechopen. doi: 10.5772/60423
- FAO (2020) The State of World Fisheries and Aquaculture 2020: Sustainability in Action. Rome. doi: 10.4060/ca9229en
- Farook MA, Mohamed HSM, Tariq N, Shariq K, Ahmed IA (2019) Giant freshwater prawn, *Macrobrachium rosenbergii* (De Man 1879): A review. Int J Res Anal Rev 6:571-584. Corpus ID: 203691772
- Gao T, Ying Y, Yang Q, Song N, Xiao Y (2020) The mitochondrial markers provide new insight into the population demographic history of *Coilia nasus* with two ecotypes (anadroumous and freshwater). Front Mar Sci 7:576161. doi: 10.3389/fmars.2020.576161
- Green MR, Sambrook J (2012) Molecular Cloning: A Laboratory Manual. 4rd Edition. Cold Springs Harbor Laboratory Press, New York
- In VV, O'Connor W, Sang VV, Van PT, Knibb W (2017) Resolution of the controversial relationship between Pacific and Portuguese oysters internationally and in Vietnam. Aquacult 473:389-399. doi: 10.1016/j.aquaculture.2017.03.004
- KKP (2014) Keputusan Menteri Kelautan dan Perikanan Republik Indonesia. Nomor 23/KEPMEN-KP/2014 Tentang Pelepasan Udang Galah GI Macro II. Kementerian Kelautan dan Perikanan,Jakarta
- KKP (2015) Keputusan Menteri Kelautan dan Perikanan Republik Indonesia Nomor 25/KEPMEN-KP/2015 Tentang Pelepasan Udang Galah SIRATU. Kementerian Kelautan dan Perikanan,Jakarta
- Li H, Kong J, Xie R, Yu W, Chen A (2022) Comparative rapid identification of Salmo salar, *Oncorhynchus mykiss*, and *Oncorhynchus keta* components based on loop-mediated isothermal amplification and quantitative

polimerase chain reaction. Aquaculture 550:737835. doi: 10.1016/j.aquaculture.2021.737835

- Li H, Yang M, Chen G, Wu Y, Xiang Y, Zhu H, Ma K, Ibrahim S, Yang G, Tang Q (2021) The complete mitogenome of giant freshwater prawn (*Macrobrachium rosenbergii*) from two different selective breeding populations in China. Mitochondrial DNA B Resour 6:1984-1986. doi: 10.1080/23802359.2021.1938720
- Li L, Lin H, Tang W, Liu D, Bao B, Yang J (2017) Population genetic structure in wild and aquaculture populations of *Hemibarbus maculates* inferred from microsatellites markers. Aquac Fish 2:78-83. doi:

10.1016/j.aaf.2017.03.004

- Li X, Deng Y, Yang K, Gan W, Zeng R, Deng L, Song Z (2016) Genetic diversity and structure analysis of *Percocypris pingi* (Cypriniformes: Cyprinidae): Implications for conservation and hatchery release in the Yalong River. PLoS One 11:e0166769. doi: 10.1371/journal.pone.0166769
- Li Y, Song J, Shen X, Cai Y, Cheng H, Zhang X, Yan B, Chu KH (2019) The first mitochondrial genome of Macrobrachium *rosenbergii* from China: Phylogeny gene and rearrangement within Caridea. Mitochondrial DNA Part В 4:134-136. doi: 10.1080/23802359.2018.1540262
- Liu MY, Cai YX, Tzeng CS (2007) Molecular systematics of the freshwater prawn genus *Macrobrachium* Bate, 1868 (Crustacea: Decapoda: Palaemonidae) inferred from mtDNA sequences, with emphasis on East Asian Species. Zool Stud 46:272-289. Corpus ID: 44152737
- Ma K, Feng J, Lin J, Li J (2011) The complete mitochondrial genome of *Macrobrachium nipponense*. Gene 487:160-
- 165. doi: 10.1016/j.gene.2011.07.017 Maidin MSR, Anton A, Yong ASK, Chin GJWL (2017) Mitochondrial COI gene sequence of giant freshwater prawn, *Macrobrachium rosenbergii*: An assessment of a community-based stock enhancement programme in

Petagas River, Sabah, Malaysia. Int J Fish Aquat Stud 5:518-526. Corpus ID: 62817893

- Mastrochirico-Filho VA, del Pazo F, Hata ME, Villanova GV, Foresti F, Vera M, Martínez P, Porto-Foresti F, Hashimoto DT (2019) Assessing genetic diversity for a pre-breeding program in *Piaractus mesopotamicus* by SNPs and SSRs. Genes 10:668. doi: 10.3390/genes10090668
- Nei M (1978) Estimation of average heterozygosity and genetic distance from a small number of individuals. Genetics 89:583-590. doi: 10.1093/genetics/89.3.583
- Nguyen Thanh H, Liu Q, Zhao L, Zhang H, Liu J, Nguyen Hai D (2015) Genetic diversity of cultured populations of giant freshwater prawn (*Macrobrachium rosenbergii*) in China using mtDNA COI and 16S rDNA markers. Biochem Syst Ecol 62:261-269. doi: 10.1016/j.bse.2015.09.011
- Novelo ND, Gomelsky B, Coyle SD, Kramer AG (2021) Evaluation of growth, sex (male proportion; sexual dimorphism), and color segregation in four cross combinations of different strains of XX female and YY male Nile Tilapia. J World Aquacult Soc 52:445-456. doi: 10.1111/jwas.12742
- Sunarma A, Carman O, Zairin Jr M, Alimuddin A (2016) Interpopulation crossbreeding of farmed and wild African catfish *Clarias gariepinus* (Burchell 1822) in Indonesia at the nursing stage. Aquat Living Resour 29:303. doi: 10.1051/alr/2016026
- Tave D (1993) Genetics for Fish Hatchery Managers. Second Edition. Springer, New Work. Corpus ID: 83320833
- Wu L, Yang J (2012) Identifications of captive and wild Tilapia species existing in Hawaii by mitochondrial DNA control region sequence. PLoS One 7:e51731. doi: 10.1371/journal.pone.0051731
- Zafar M, Soomro MH, Daudpota AM, Memon AJ, Ishaqi AM (2015) Effect of different salinities on survival of freshwater prawn (*Macrobrachium rosenbergii*) larvae at seed production unit Hawksbay Karachi - Pakistan. Int J Interdiscip Multidiscip Stud 2:165-169

- Zeng Q, Sun C, Dong J, Tian Y, Ye X (2017) Comparison of the crossbreeding effects of three Mandarin fish populations and analyses of the microsatellite loci associated with the growth traits of F1 progenies. Int J Aquacult Fish Sci 3:035-041. doi: 10.17352/2455-8400.000026
- Zhang WZ, Xiong XM, Zhang XJ, Wan SM, Guan NN, Nie CH, Zhao BW, Hsiao CD, Wang WM, Gao ZX (2016)

Mitochondrial genome variation after hybridization and differences in the first and second generation hybrids of bream fishes. PLoS One 11:e0158915. doi: 10.1371/journal.pone.0158915

Zhang X, Wu W, Li L, Ma X, Chen J (2013) Genetic variation and relationships of seven sturgeon species and ten interspecific hybrids. Genet Sel Evol 45:21. doi: 10.1186/1297-9686-45-21